

ISOLATION AND IDENTIFICATION OF SOME PHYTOCHEMICAL COMPOUNDS FROM DIFFERENT PARTS OF *IRIS NIGRICANS*

Sajee' Huwaitat

Ekbal Al-Khateeb

Samira Finjan

Pharmaceutical Pharmacy department, Faculty of Pharmacy & Medical Sciences,
Al-Ahliyya Amman University, Jordan

Abstract

Iris genus (Iridaceae) is an elegant endemic plant species, and considered threatened to Jordan due to habitat degradation. Rhizomes of some *Iris* species contain flavonoids such as iridin, iriline A, irisone B. *Iris* leaves contain many flavonoids such as irilin, iristectorigenins, tectorigenin, and coumaronochromones. *Iris* flowers contain also flavonoids and proanthocyanidin while *Iris nigricans* have no previous studies conducted on leaves and flowers. In this study some phytochemical compounds were separated from different parts of *Iris nigricans* (rhizomes, leaves and flowers) by methanol. Isolation is carried out by using chromatographic technique and identified by ferric chloride reagent and UV light λ_{max} 254-365nm. The phenolic composition of methanolic extracts of *Iris nigricans* obtained from rhizomes, leaves and flowers were elucidated by MS, $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$.

Keywords: Methanolic extracts of *Iris nigricans* (different parts), chromatographic technique, MS, $^1\text{H-NMR}$, $^{13}\text{C-NMR}$, mangleferin, iristectoreginin A, proanthocyanidin

Footnotes: MS= Mass Spectrometry, $^1\text{H-NMR}$ = Proton Nuclear Magnetic Resonance, $^{13}\text{C-NMR}$ = Carbon 13 Nuclear Magnetic Resonance.

Introduction

Medicinal plants have an essential role in the drug discovery and many modern drugs have their origin in traditional medicines of different cultures. Although the synthetic and combinatorial chemistry as well as molecular modeling are beneficial, medicinal plants

remain an essential source of new drugs, new drug leads and new chemical entities (NCEs). (Rana Abu-Dahab, et al., 2007)

Iris species which have different chemical constituents (glycosides, phenolic compounds and xanthenes) which related to different biological activities such as anti-inflammatory, antioxidant activity, antiseptic, pain relief, antibacterial and antifungal. (Das DK, et al., 1999, Sajee' Huwaitat, et al., 2013)

Iris leaves contain many isoflavones particularly iristectorigenin A which has different biological activities such as estrogenic activity. (Eva Miadokova', 2009)

Other constituent like xanthone is found in Iris genus used as an insecticide (Jian Zhao, et al., 2005). Proanthocyanidins compounds are found in this genus which related to different biological activities. (Das DK, et al., 1999, Sajee' Huwaitat, et al., 2013)

Methodology

Plant Material

The rhizomes, leaves and flowers of *Iris nigricans* grown in Jordan were collected during the months of March and April (2012). The rhizomes and leaves were dried at room temperature in the shade for about 15 days and then weighed. The flowers were cut into small pieces and weighed.

Extraction Methods

Extraction of rhizomes

Dried rhizomes (12.800 gm) were grinded into powder by a mixer followed by extraction with 80% methanol in soxhlet apparatus for 48 hours at temperature between 60 - 70°C. The evaporated methanolic extract (1.629gm) was collected in amber glass containers to avoid any oxidation. (Nighat Nazir, et al., 2008, Syeda F., et al., 2009).

Extraction of leaves and flowers

Fresh leaves (12.200gm) and fresh flowers (10gm) were macerated in 70% v/v and 95% v/ of methanol for one week in refrigerator. The evaporated extracts were kept in amber glass containers to avoid any oxidation. (V. H. Booth, et al., 1960, E. Nicole Bridgers, et al., 2010).

Chromatographic technique

All extracts (rhizomes, leaves and flowers) were subjected to thin-layer chromatography using the mobile phases (chloroform 68 ml: methanol 32 ml), (chloroform 68 ml: methanol 32 ml) and (chloroform 50 ml: methanol 50 ml) respectively. All the chromatograms were visualized by UV light (λ_{254} nm – λ_{365} nm) and then spraying with ferric chloride solution.

Isolation and Identification Methods

One band from each extracts of *Iris nigricans* (rhizomes, leaves and flowers) were isolated by using preparative TLC (20 x 20 cm) coated with silica gel GF₂₅₄ layers of 1 mm thickness and using the same mobile phases as mentioned before. These bands were selected according to the presence of orange color and clear fluorescence under UV light and to the width of the bands. The isolated compounds out of the silica gel were subjected for further spectral analysis (MS, ¹H-NMR and ¹³C-NMR).

Results and Discussion

Extraction Method

All the extracts from the rhizomes, leaves and flowers were gave dark blue color due to the formation of colored complex Fe(OAr)₃. (Ranju Pal, et al., 2012).

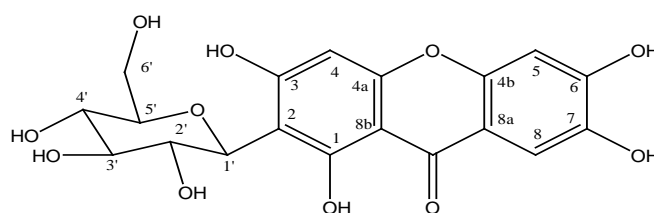
According to the Preparative Thin-Layer Chromatography of the rhizomes, among the bands one yellow band in daylight gave the clear fluorescent band and named by band 1 for rhizomes which subjected for further spectral analysis (MS, ¹H-NMR and ¹³C-NMR) and gave the following results.

EI-mass spectrum of band 1 from yellow band 2 of rhizomes was measured on high resolution mass spectrometer using ESI technique (electron spray ionization) and methanol and chloroform as solvent. The spectrum showed the following bands at *m/z*: 422 (M⁺), 423 (M⁺ +1), 424 (M⁺ +2) due to the existence C isotope.

¹H-NMR spectrum (MeOH-d₄, 500 Hz) of this fraction showed the appearance of the following bands (δ ppm): 7.32 (1H,s, H-8); 6.39 (1H,s, H-5); 6.19(1H,S, H-4); (5.00-4.96) (free OH_s exchangeable with D₂O); (4.15-3.95) (2H, m, H-1' and H-5'); (3.75-3.65) (3H, m, H-2', H-3', H-4'); (3.55- 3.45) (2H, m, H-6').

¹³C-NMR spectrum (MeOH-d₄, 500 Hz) showed the following chemical shifts δ (ppm): 182.20 (C=O); 162.10 (C-3); 160.10 (C-1); 155.70 (C-4a); 150.70 (C-6); 149.70 (C-4b); 140.10 (C-7); 117.20 (C-8a); 115.00 (C-8); 106.30 (C-2); 105.40 (C-5), 102.40 (C-8b); 97.30 (C-4); 85.00 (C-5'); 80.70 (C-3'); 77.20 (C-1'); 72.90 (C-2'); 70.50 (C-4'); 65.30 (C-6').

The above spectral data (MS, ¹H-NMR and ¹³C-NMR) for this band is consistent with the structure of mangiferin presented below. (Nathalie Wauthoz, et al., 2007 and Watson, 2001)



Mangiferin (C₂₀H₂₂O₁₀)

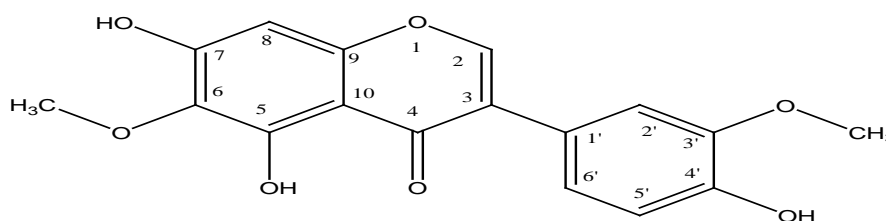
According to the Preparative Thin-Layer Chromatography of the leaves, orange band in daylight and under UV light was appeared clearly and then subjected to spectral analysis by MS, $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$. Orange band gave the following results.

EI-mass spectrum of orange band was measured on high resolution mass spectrometer using ESI technique (electron spray ionization). The spectrum showed peaks at m/z : 330 (M^+), 331 ($\text{M}^+ + 1$) and 332 ($\text{M}^+ + 2$) due to the existence C isotope.

$^1\text{H-NMR}$ spectrum (MeOH-d_4 , 500 Hz) of this fraction showed the appearance of the following bands δ (ppm): 7.75 (1H, m, H-2); 7.15 (1H, m, H-5'); 6.95 (1H, m, H-6'); 6.55 (1H, m, H-2'); 6.25 (1H, s, H-8); 5.5- 5.25 (3H, br.s, free OHs exchangeable with D_2O); 3.85 (6H, s, OCH_3 -6 and OCH_3 -3').

$^{13}\text{C-NMR}$ using MeOH-d_4 as solvent and high resolution 500 Hz. Spectrum showed the following chemical shifts δ (ppm): 175.40 (C-4); 153.60 (C-9); 153.20 (C-2); 152.40 (C-7); 147.20 (C-5); 146.50 (C-4'); 137.10 (C-6 and C-3'); 126.60 (C-1'); 123.50 (C-3); 120.40 (C-6'); 117.20 (C-2'); 113.60 (C-5'); 106.50 (C-10); 99.00 (C-8); 56.50 (OCH_3 -6 and OCH_3 -3').

These spectral data (MS, $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$) for this fraction are consistent with the structure of iristectorigenin A presented below. (Le Minh Ha, et al., 2009 and Watson, 2001)



Irictectorigenin A ($\text{C}_{17}\text{H}_{14}\text{O}_7$)

The results for the Preparative Thin-Layer Chromatography of the flowers, band 1 out of several bands was chosen for clear fluorescent band under UV light then this band was subjected to different spectral analysis including MS, $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$, the results were illustrated.

EI-mass spectrum of band 1 from total flowers extract was measured on high resolution mass spectrometer using ESI technique (electron spray ionization).

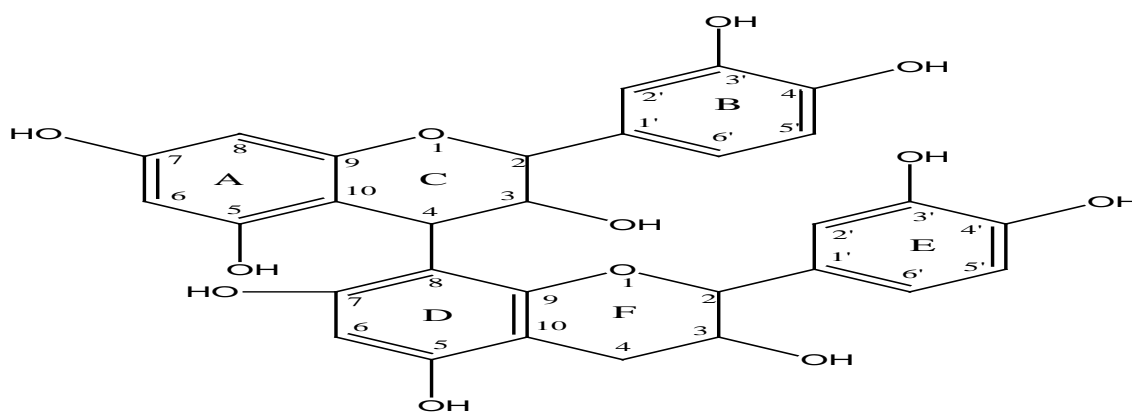
The spectrum showed the following peaks at m/z : 578 (M^+); 579 ($\text{M}^+ + 1$) and 580 ($\text{M}^+ + 2$) due to the existence C isotope.

$^1\text{H-NMR}$ spectrum (MeOH-d_4 , 500 Hz) of this fraction showed the appearance of the following bands δ (ppm): 7.65-7.55 (6H, m, H-2', H-5', H-6' in rings B and E); 7.25-7.15 (3H,

m, H-6 and H-8 in ring A and H-6' in ring D) ; 5.35- 5.15 (10H, br.s, free OHs exchangeable with D₂O); 4.95- 4.85 (2H, m, H-2 in rings C and F); 4.45- 4.25 (2H, m, H-3 protons in rings C and F); 2.95- 2.85 (1H, m, H-4 in ring C); 2.75-2.55 (2H, m, H-4 in ring F).

¹³C-NMR spectrum (MeOH-d₄, 500 Hz) showed the appearance of the following chemical shifts δ (ppm): 158.80 (C-7 in ring A); 157.40 (C-5 in ring A); 155.70 (C-5 in ring D); 154.80 (C-9 in ring A and C-7, C-9 in ring D); 147.40 (C-4' in ring B and ring E); 144.60 (C-3' in ring B and ring E); 132.60 (C-1' in ring B and ring E); 122.20 (C-6' in ring B and ring E); 117.40 (C-5' in ring B and ring E); 115.20 (C-2' in ring B and ring E); 82.40 (C-2 in ring C); 79.50 (C-2 in ring F); 72.70 (C-3 in ring C); 67.80 (C-3 in ring F); 29.20 (C-4 in ring C) and 26.90 (C-4 in ring F).

These spectral data (MS, ¹H-NMR and ¹³C-NMR) for this fraction are consistent with the structure of dimeric proanthocyanidin presented below. (Kohei Kamiya, et al., 2001 and Watson, 2001).



Proanthocyanidin (C₃₀H₂₆O₁₂)

Conclusion

Since there is no previous studies on the chemical constituents of the leaves and flowers of *Iris nigricans* grown in Jordan, this research showed the isolation and identification of some of phenolic compounds (glycosides, flavonoids and xanthones). Mangiferin was elucidated in rhizomes, the isoflavonoid iristectorigenin A was documented in leaves and proanthocyanidin was determined in flowers.

References:

- Das DK, Sato M, and Ray PS, Cardioprotection of red wine: role of polyphenolic antioxidants, *Drugs Exp Clin Res*, 1999, 25:115-20.
- David G. Watson, pharmaceutical analysis, 2001, Pages 149-170.

E. Nicole Bridgers, Mari S. Chinn and Van-Den Truong, Extraction of anthocyanins from industrial purple-fleshed sweetpotatoes and enzymatic hydrolysis of residues for fermentable sugars, *Industrial Crops and Products*, 2010, Volume 32, Pages 613–620.

Eva Miadokova', Isoflavonoids – an overview of their biological activities and potential health benefits, *InterdiscToxicol.*; 2009, Volume 2, No. 4, Pages 211–218.

Kohei Kamiya, Chiharu Watanabe, Hanani Endang, Mansur Umar and Toshiko Satake, Studies on the Constituents of Bark of *Parameria laevigata* MOLDENKE, *Chem. Pharm. Bull.* 2001, Volume 49, No. 5, Pages 551—557.

Le Minh Ha, Pan Van Kiem and Natalya Khripach, Chemical constituents of *Belamcanda Chinensis* (L.) DC, *Journal of Chemistry*, 2009, Volume 47, No. 5, Pages 623 – 627.

Nathalie Wauthoz, Aliou Balde, Elhadj Saïdou Balde, Marc Van Damme and Pierre Duez, Ethnopharmacology of *Mangifera indica* L. bark and pharmacological studies of its main C-Glucosylxanthone, Mangiferin, *International Journal of Biomedical and Pharmaceutical Sciences*, 2007, Pages 113-119.

Nighat Nazir, Surrinder Koul and Mushtaq Ahmad Qurishi, New isoflavones from *Iris kashmiriana*, *Journal of Asian Natural Products Research*, 2008, Volume 10, No.12, Pages 1137–1141.

Rana Abu-Dahab, and Fatma Afifi, Antiproliferative activity of selected medicinal plants of Jordan against a breast adenocarcinoma cell line (MCF7), *Scientia Pharmaceutica*, 2007, Volume75, Pages121-136.

Ranju Pal, Kundlik Girhepunje, Ashutosh Upadhayay and Thirumoorthy N., Antioxidant and free radical scavenging activity of ethanolic extract of the root of *Morinda citrifolia* (Rubiaceae), *African Journal of Pharmacy and Pharmacology*, 2012, Volume 6, No.5, Pages 278-282.

Sajee' Huwaitat, Ekbal AL-khateeb, Samira Finjan and Anwar Maraqa, Antioxidant and antimicrobial activities of *Iris nigricans* methanolic extracts containing phenolic compounds, *European Scientific Journal*, 2013, Volume 9, No. 1, Pages 1-10.

Syeda F. Asghar, Shahid Aziz and Habib-ur-Rehman, Secondary Metabolites Isolated from *Iris germanica*, *Records of Natural Products*, 2009, Volume 3, No.3, Pages139-152.

V. H. Booth and B. J. Constable, Stability of ascorbic acid in extracts of *Iris* leaves, *Biochemical Journal*, 1960, Volume76, Page 206.