

ESJ Natural/Life/Medical Sciences

Comparative Effects of Some Botanical Extracts and Chemicals in Controlling the Red Spider Mite *Tetranychus evansi* Baker & Pritchard on Solanaceous Crops

G.Y. Azandémè-Hounmalon

Enseignant-Chercheur à l'Ecole d'Horticulture et d'Aménagement des Espaces Verts (EHAEV) de l'Université Nationale d'Agriculture (UNA), République du Bénin

A. Onzo

Enseignant-Chercheur à la Faculté d'Agronomie. Laboratoire de Phytotechnie, d'Amélioration et de Protection des Plantes (LaPAPP); Université de Parakou, République du Bénin

A. Adandonon

Enseignant-Chercheur à l'Ecole de Gestion et de Production Végétale et Semencière (EGPVS) de l'Université Nationale d'Agriculture (UNA)

J. Houedenou

Master en Production végétale à l'Ecole de Gestion et de Production Végétale et Semencière (EGPVS) de l'Université Nationale d'Agriculture (UNA)

R. Djossou

Doctorant à l'Ecole Doctorale des Sciences Agronomiques et de l'Eau (EDSAE) de l'Université Nationale d'Agriculture (UNA)

D. Gnanvossou

M. Tamò

Chercheurs à International Institute of Tropical Agriculture (IITA), République du Bénin

Doi:10.19044/esj.2022.v18n33p101

Submitted: 08 July 2022 Accepted: 12 October 2022 Published: 31 October 2022 Copyright 2022 Author(s) Under Creative Commons BY-NC-ND 4.0 OPEN ACCESS

Cite As:

Azandémè-Hounmalon G.Y., Onzo A., Adandonon A., Houedenou J., Djossou R., Gnanvossou D. & Tamò M. (2022) *Comparative Effects of Some Botanical Extracts and Chemicals in Controlling the Red Spider Mite Tetranychus evansi Baker & Pritchard on Solanaceous Crops.* European Scientific Journal, ESJ, 18 (33), 101. <u>https://doi.org/10.19044/esj.2022.v18n33p101</u>

Abstract

The tomato red spider mite, Tetranychus evansi, is an invasive pest reported on solanaceous crops in Benin around 2008, causing heavy economic damage. The control of this mite by farmers is mainly done through intensive applications of chemical pesticides that are not always effective. In the present study, we evaluated in a laboratory, at IITA-Benin, the effects of two botanical insecticides: the Cashew Nut Shell Liquid (CNSL) and Neem oil. This is together with Acarius 18 EC, a chemical acaricide generally used by growers on eggs and adult females T. evansi. The half recommended dose (0.5 l/ha), the recommended dose (1 l/ha), and the double recommended dose (2 l/ha) of Acarius and Neem oil as well as solutions at 1%, 2%, 3%, 4% and 5% of CNSL were tested. Mean egg hatching rates varied significantly among treatments, ranging from $0.00 \pm 0.00\%$ (Neem oil and CNSL) (respectively at half recommended dose and 4%) to 100% (Control). Mortality of adult female T. evansi also differed significantly among treatments (P < 0.0001), ranging from $22.00 \pm 4.20\%$ to 100%. The highest mortality rates were recorded with Neem oil at any doses and with CNSL at 4%, whereas the lowest rate was recorded with the control treatment. Fecundity of pesticide-treated females T. evansi and proportion of eggs that hatched revealed significant differences among all doses of products (P < 0.0001). It appears from this study that even the half recommended dose of Neem oil and the CNSL at 4% were very effective on suppressing T. evansi populations and should, therefore, be subject to further studies to test their compatibility with natural enemies, and to determine strategies for their efficient applications in greenhouse and under field conditions.

Keywords: Mite pests of tomato, Cashew Nut Shell Liquid (CNSL), Neem oil, *Solanum lycopersicon*, biopesticide

Introduction

The tomato red spider mite, *Tetranychus evansi* Baker and Pritchard (Acari: Tetranychidae), is an invasive pest of tomato and other solanaceous crops in several African, American, Asian, and European countries, with the potential of causing up to 90% yield reduction (Sarr *et al.*, 2002; Saunyama & Knapp, 2003; Navajas *et al.*, 2012). In the Republic of Benin (West Africa), *T. evansi* has become, since 2008, the key mite pest of first importance not only of tomato (*Solanum lycopersicon* L.), but also of other solanaceous crops such as the African eggplant (*Solanum macrocarpon* L.) (Azandémè-Hounmalon *et al.*, 2014, 2015). This mite is characterized by its high reproductive capacity, which leads to high population levels in a short time resulting in an important economic damage (Moraes & McMurtry, 1986). It could destroy a whole solanaceous field in less than one week if no adequate

treatment is applied (Azandémè-Hounmalon *et al.*, 2014). Besides its invasive nature, recent reports indicate that *T. evansi* may be displacing native spider mite species, hence posing new pest management challenges (Ferragut *et al.*, 2013).

While there are efforts to control T. evansi using cultural practices (Saunyama & Knapp, 2003; Savi et al., 2019ab; Djossou et al., 2020; Savi et al., 2021), application of chemical pesticides remains the method preferred by farmers (Toroitich et al., 2014; Azandémè-Hounmalon et al., 2015). However, previous studies have shown that chemical applications can entail the risk of resistance development among insect and mite populations because of longterm exposure to pesticides (Blair, 1989; Tsagkarakou et al., 2002; Nyoni et al., 2011). Thus, this result to the recrudescence of the pest as these pesticides had lost their efficacy (Copping & Menn, 2000; Marcié & Medo, 2014; Reddy, 2016). As a result, there is an urge to search for alternative methods that can efficiently control this mite pest without harming the environment. Fortunately, biopesticides such as plant extracts and essential oils have been acknowledged to be effective while being biodegradable, with no hazardous effects on the environment (Sanon et al., 2005; Rochefort et al., 2006; Salma & Jogen, 2011; Sarwar, 2015). As a consequence, several studies have evaluated the efficacy of plant extracts in the control of agricultural and crop pests including vegetable mite pests (Adango et al., 2020) and insect pests (Gnago et al., 2010; Fayalo et al., 2014; Sane et al., 2018), as well as other arthropod pests associated with bees and bee products (Gbedomon et al., 2012).

In Africa, several plant species are known and used for their biocidal activities (toxic, repellent, anti-appetite) against pests (Baidoo *et al.*, 2012). Neem (*Azadirachta indica* A. Juss) is one of such plant whose extracts (leaf, seed oil etc..) are being extensively used as an alternative to chemical pesticides in managing insect and mite pests (Fayalo *et al.*, 2014; Traoré *et al.*, 2019a; Adango *et al.*, 2020). The major active ingredient of Neem extracts is the azadirachtin that can cause, on arthropods, several negative effects such as feeding inhibition, repellency, fertility and fecundity reduction, behavioral changes, and increased mortality (Schmutterer, 1990; Dimetry *et al.*, 1993; Mordue & Nisbet, 2000; Musabyimana *et al.*, 2001). The main advantages of using neem extracts are their insecticidal and acaricidal activities, their low toxicity towards mammals and birds, and their fast degradation in the soil and animals (Isman, 2006; Normas, 2008).

The cashew tree (*Anacardium occidentale* L.) is a multipurpose tree. In Benin, cashew is the second largest export crop after cotton. In 2013, Benin was ranked 5^{th} exporter of raw cashew nuts among the main world exporters with a production of 180.000 tons / year (FAO, 2013). However, in spite of such a high production, some of the products and by-products of cashew trees

are often thrown away, although some of them could have biocidal properties that could be considered in the control of crop pests. In their study, Chabi *et al.* (2014) have shown that leaves and bark of cashew tree have fungicidal and bactericidal properties. Other studies have shown that Cashew Nut Shell Liquid (CNSL: the balm derived from shell) have interesting biocidal properties against pests (Cavalcante *et al.*, 2003; Chabi *et al.*, 2013; 2014).

To control mite and other insect pests, Acarius 18EC (Abamectin 18g / L, EC) is the chemical insecticide-acaricide widely used by farmers in major vegetable production in Benin (Azandémè-Hounmalon GY unpublished). The active ingredient of Acarius 18 EC is Abamectin that has been enriched with oil to facilitate its penetration into the plant tissue. It acts by ingestion and, to a lesser extent, by contact on mobile forms of mites and on biting-sucking insects.

Whereas Neem oil have been shown to cause deleterious effects on some mite species (Adango *et al.*, 2020; Azandémè-Hounmalon *et al.*, in press), no studies have yet been carried out to determine the acaricidal properties of the CNSL.

Therefore, this paper focuses on evaluating the efficacy of Neem oil and CNSL (two botanical insecticides) in comparison with Acarius 18 EC (chemical insecticide-acaricide) in controlling the tomato red spider mite *T. evansi*, a first step for their possible use as alternatives to chemical pesticides on vegetable farms.

2. Methods

2.1 Study Site

The study was conducted in a laboratory at the International Institute of Tropical Agriculture (IITA), Benin-Station, located at 12 km NW Cotonou (6°25'N; 2°19'E; 15 m asl). The experimental conditions were 25 ± 2 °C, 60–70% RH and 12:12 h (L: D).

2.2 Plants and Mites Rearing

The potted tomato plants used in the study were kept on metallic benches in a screenhouse for about 45 days until they bear at least four completely developed leaves. The leaves were subsequently picked, either for producing leaf discs or for rearing *T. evansi*.

The population of *T. evansi* used in this study was originally collected from a vegetable farm at Sèmè-Kpodji in the Southern-Benin (6°22'N; 2°37E) in March 2016 and maintained in a screenhouse at IITA-Benin (27 \pm 1 °C, 70–80% RH and 12:12 h (L:D) photoperiod) on nightshade (*Solanum macrocarpon*) or tomato plants.

2.3. Pesticides Tested

The pesticides used in this study included the Cashew Nut Shell Liquid (CNSL) and Neem oil, two biopesticides, and Acarius 18 EC, which is a chemical acaricide (Abamectin 18g/L, EC) commonly used by growers on vegetable farms to control arthropod pests.

Neem oil is obtained from *Azadirachta indica* A. Juss. (Meliaceae) seeds and is used as an insecticide whose active ingredient is Azadirachtin. It is a 100% natural oil that is non-toxic to humans and animals. It does not act by contact but prevents insects from feeding thereby disturbing their development and their ability to oviposit (Karnavar, 1987; Belanger & Musabyimanan, 2005). The Neem oil used in this study was purchased from Biophyto at Allada in the Republic of Benin.

The chemical pesticide Acarius 18 EC (Abamectin 18g/L, EC), miticide-insecticide, is manufactured by Savana Www.Savanna-France.Com. The application dose recommended is 0.5 l/ha. Abamectin acts by ingestion and, to a lesser extent, by contact on biting-sucking mites and insects (RECA, 2013; http://terra.mg/agricom/agriculture/fr/produit/acarius-018-ec/, June 2022).

The CNSL was obtained from the Nutrition Department at the Faculté des Sciences Agronomiques (FSA) of the Université d'Abomey-Calavi (UAC) in Benin. The different concentrations of CNSL tested in this study were based on the method reported by Kerala Agricultural University (2009).

2.4 Experimental Design

The experiment consists of the application of the two botanical insecticides (i.e., Neem oil and CNSL), the chemical acaricide-insecticide (i.e., Acarius 18EC), and the control treatment (application of tap water alone). Three different doses of Neem oil and Acarius including half of recommended dose (D1), the recommended dose (D2), and the double of the recommended dose (D3) were tested.

According to the recommendations by RECA (2013), 25 ml of Acarius should be mixed with 15 liters of water while 80ml of Neem oil are requested for 16 liters of water (https://www.biophyto-benin.com). For CNSL, five solutions of the respective concentrations of 1%, 2%, 3%, 4%, and 5% were prepared in distilled water with vegetable soap (palmida) added as emulsifier. To achieve it, 50g of the soap were dissolved in 670 ml of water. Then, 330 ml of CNSL was added to the mixture which was vigorously stirred to obtain 11 of a good emulsion (i.e., the stock solution). Series of dilutions were carried out using the stock solution to obtain the different concentrations of CNSL tested (1%, 2%, 3%, 4% and 5%).

For each pesticide, the solution used in the experiments was prepared as presented in Table 1.

Table 1. Different doses used for acaricide treatment

Treatments	D1	D2	D3	1%	2%	3%	4%	5%
Acarius	0.83	1.66	3.34	-	-	-	-	-
Neem oil	2.5	5	10	-	-	-	-	-
CNSL	-	-	-	10	20	30	40	50

Doses (ml/l) of water

D1 = Half recommended dose; D2 = Recommended dose; D3 = Double recommended dose 1%, 2% 3% 4% and 5% = concentrations of CNSL

2.5 Data Collection

2.5.1 Effects on Adult Females T. evansi

Per treatment, 5 leaf discs cut from fresh *S. lycopersicum* leaves were dipped for 5 seconds into the pesticide solution tested. The leaf discs (2 cm diameter) were then placed on the lower side up on water-soaked cotton wool in Petri dishes (9 cm diameter), and it was allowed to dry for 30 minutes (Toroitich *et al.*, 2014). Thereafter, 10 adult females *T. evansi* (24 to 48h-old) were transferred from the rearing unit to each leaf disc. The monitoring started 24 h later for six consecutive days during which the daily mortality of females, the number of eggs laid, and those that hatched were recorded. Escaped mites were excluded from the analysis.

2.5.2 Effects on *T. evansi* Eggs

Per treatment, 5 leaf discs (25 mm diameter) were cut from fresh *S. lycopersicum* leaves and on each of them, 10 newly laid *T. evansi* eggs (24 h-old) were deposited. The 5 leaf discs for each treatment were then placed, lower side up, on water-soaked cotton wool in a Petri dish (9 cm diameter) and sprayed with the pesticide solution at its appropriate dose for 5 seconds. The leaf discs were then dried out at ambient air for 1h. The leaf discs contained in a Petri dish represent the 5 replicates for each treatment. The leaves were monitored daily for five consecutive days to record the number of eggs that hatched per treatment and per dose.

2.5.3 Data Analysis

The mortality data on ovicidal tests were corrected in relation to the control (water) using the Abbott's formula (Abbott, 1925), while actual uncorrected figures were used for adult mortality. Data on egg and adult mortality rates were arcsine square root-transformed (Arcsine $\sqrt{(X)}$), which is then subjected to a one-way analysis of variance (Proc GLM). In case of significant differences, treatments means were separated using the Student-Newman-Keuls test (SAS 9.1, 2009).

3. Results

3.1 Effect on Mortality of Adult Female T. evansi

The temporal evolution of different product effect on the mortality of adult female *T. evansi* is shown in Figure 1 and Table 2. The mean mortality rate of adult female *T. evansi* varied between $22.00 \pm 4.20\%$ (Control) to 100% (CNSL 4%, 5% and Neem oil). At day 1 (i.e., 24 hours) after application, the highest mortality rate of *T. evansi* females (100%) reached half recommended dose (D1) of Neem oil, 4% and 5% of CNSL. Also, 3 days after application of Acarius, this rate reached the double recommended dose (D3). At the end of the observation period, the analysis of variance showed significant differences in the effects of treatments on the mortality of adult female *T. evansi* (df = 11; F = 39.82; P < 0.0001; Table 2). The Student-Newman Keul test revealed that all the doses of Neem oil as well as the 4% and 5% of CNSL solutions had statistically similar effects by inducing the highest mortality of adult female *T. evansi* (Table 2).

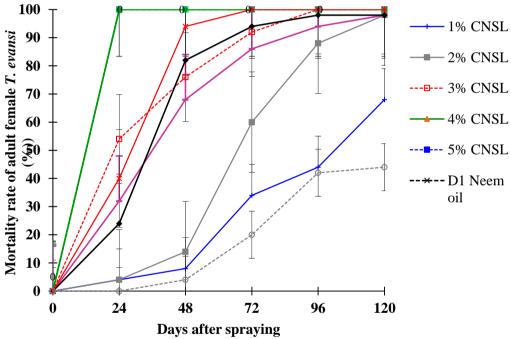


Figure 1. Adult mortality of *T. evansi* at 1%, 2%, 3%, 4%, 5% of CNSL, at half recommended dose D1, recommended dose D2 and double recommended dose D3 of different pesticide products

Treatments	Ν	Mean \pm SE	Df	F	Р
CNSL 1%	25	$31.60\pm5.67a$			
CNSL 2%	25	$52.80\pm8.13b$			
CNSL 3%	25	$84.40 \pm 4.69 cd$			
CNSL 4%	25	$100.00\pm0.00d$			
CNSL 5%	25	$100.00\pm0.00d$	11	39.82	< 0.0001
Acarius D1	25	$75.60\pm5.41c$			
Acarius D2	25	$79.20\pm5.91c$			
Acarius D3	25	$86.80 \pm 5.43 cd$			
Neem oil D1	25	$100.00\pm0.00d$			
Neem oil D2	25	$100.00\pm0.00d$			
Neem oil D3	25	$100.00\pm0.00\text{d}$			
Control	25	$22.00\pm4.20a$			

Table 2. Effects of treatments on mortality of adult female *T. evansi*

Values represent Means \pm SE. Means followed by the same letter in a column are not significantly different for the performance measurement (SNK test, P < 0.05); D1 = Half recommended dose; D2 = Recommended dose; D3 = Double recommended dose; 1%, 2%, 3%, 4% and 5% = concentrations of CNSL N= Number of observations

3.2 Effects on Viability of *T. evansi* Eggs

Among all the three pesticides tested (all doses pooled together), the average hatching rates of *T. evansi* eggs ranged from 0% (Neem oil, CNSL 4% and 5%) to 100% (Control) (Figure 2). Results of the analysis of variance showed significant differences in the viability of *T. evansi* eggs among all the treatments (df = 11; F = 46.7; P < 0.0001). However, no eggs hatching was recorded with Neem oil and CNSL (4% and 5%) applications (Table 3).

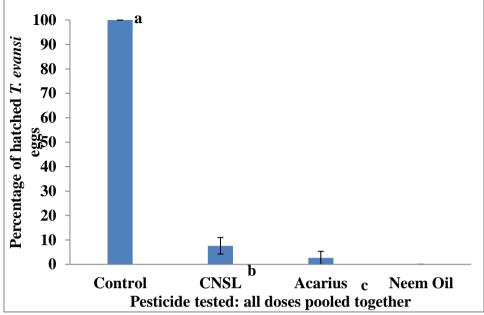


Figure 2. Eggs hatching of *T. evansi* after application of pesticide products Columns followed by the different letters indicate significant differences (SNK test, P<0.05)

Treatments	Ν	$Mean \pm SE$	Df	F	P
CNSL1%	5	$32.00 \pm 11.57 b$			
CNSL2%	5	$2.00 \pm 1.99 aa$			
CNSL3%	5	$4.00\pm4.02a$			
CNSL4%	5	$0.00\pm0.00a$			
CNSL5%	5	$0.00\pm0.00a$			
Acarius D1	5	$8.00\pm7.99a$			
Acarius D2	5	$0.00\pm0.00a$	11	46.7	< 0.0001
Acarius D3	5	$0.00\pm0.00a$			
Neem oil D1	5	$0.00\pm0.00a$			
Neem oil D2	5	$0.00\pm0.00a$			
Neem oil D3	5	$0.00\pm0.00a$			
Control	5	$100 \pm 0.00c$			

Table 3. Within-treatments effects of doses on hatc	chability of T. evansi eggs
---	-----------------------------

Values represent Means \pm SE. Means followed by the same letter in a column are not significantly different for the performance measurement (SNK test, P < 0.05); D1 = Half recommended dose; D2 = Recommended dose; D3 = Double recommended dose; 1%, 2%, 3%, 4% and 5% = concentrations of CNSL N= Number of observations

3.3 Fecundity of Adult Females *T. evansi* that Survived Pesticide Applications

The mean number of eggs laid by an adult female *T. evansi* that survived the pesticide applications ranged between 1.33 ± 0.32 (Acarius) and 24.68 ± 3.78 (Control). The results of the analysis of variance showed significant differences among treatments (df = 6; F = 9.28; P < 0.0001) (Table 4). However, there were no surviving *T. evansi* females with 4%, 5% of CNSL, and the Neem oil applications. For all the products, the lowest fecundity rate for the surviving *T. evansi* females was recorded with Acarius (1.33 ± 0.32).

Treatments	Ν	Mean \pm SE	Df	F	Р
CNSL 1%	25	$8.16 \pm 0.67 \mathrm{ac}$			
CNSL 2%	16	$9.68 \pm 1.76c$			
CNSL 3%	8	$2.50\pm0.88b$			
CNSL 4%	-	-			
CNSL 5%	-	-			
Acarius D1	6	$3.00\pm0.68ab$	6	9.28	< 0.0001
Acarius D2	8	$4.25 \pm 1.55 bc$			
Acarius D3	3	$1.33 \pm 0.32 ab$			
Neem oil D1	-	-			
Neem oil D2	-	-			
Neem oil D3	-	-			
Control	25	$24.68\pm3.77d$			

Table 4. Fecundity of adult females T. evansi after application of pesticide products

Values represent Means \pm SE. Means followed by the same letter in a column are not significantly different for the performance measurement (SNK test, P < 0.05); D1 = Half recommended dose; D2 = Recommended dose; D3 = Double recommended dose; 1%, 2%, 3%, 4% and 5% = concentrations of CNSL N= Number of observations

3.4 Viability of Eggs laid by Pesticide-treated Adult Females *T. evansi*

The hatching rate of eggs laid by pesticide-treated female *T. evansi* was significantly the lowest on CNSL 2% (0.40 ± 0.40) compared to the Control (20.00 ± 8.16). The results of the analysis of variance revealed significant differences among the different doses of products (df = 11; F = 5; P <0.0001) (Table 5). However, irrespective of the products and doses tested, none of those eggs were able to hatch.

European Scientific Journal, ESJ October 2022 edition Vol.18, No.33

Treatments	Ν	Mean ± SE	Df	F	Р
CNSL 1%	25	$6.40\pm3.36b$			
CNSL 2%	25	$0.40 \pm 0.40a$			
CNSL 3%	25	$0.8 \ 0 \pm 0.80 a$			
CNSL 4%	25	-			
CNSL 5%	25	-			
Acarius D1	25	$1.60 \pm 1.60 ab$	11	5.0	< 0.0001
Acarius D2	25	-			
Acarius D3	25	-			
Neem oil D1	25	-			
Neem oil D2	25	-			
Neem oil D3	25	-			
Control	25	$20.00\pm8.16c$			

Table 5. Viability of T. evansi eggs after application of pesticide products

Values represent Means \pm SE. Means followed by the same letter in a column are not significantly different for the performance measurement (SNK test, P < 0.05); D1 = Half recommended dose; D2 = Recommended dose; D3 = Double recommended dose; 1%, 2%, 3%, 4% and 5% = concentrations of CNSL N= Number of observations

Discussion

The use of pesticide plants extracts is now emerging as one of the important tools in crop protection, which preserve the environment. Many of such plants are known and used for their biocidal activities (toxic, repellent, anti-appetizing) against a wide range of pest (Yarou, 2017).

The results accrued from this study showed that the biopesticides Neem oil (half recommended dose) and CNSL (4% and 5% of concentration) exerted the most significant effects against T. evansi by inducing higher mortality (100%) and by reducing or neutralizing the fecundity and viability of their eggs compared with Acarius. It appears that Neem oil and CNSL have had ovicidal effects on T. evansi, and this could be explained by T. evansi egg having a slender chorion with airfoils (small holes related to embryonic respiration), which certainly facilitate its penetration by the toxic compounds (Bruce et al., 2004). Similar results were reported by Asogwa et al. (2007), who showed that various concentrations of CNSL solutions (i.e., 6%, 8% and 10%) caused 100% mortality in termite populations. Kpoviessi et al. (2017a) reported that the CNSL can significantly reduce aphids, thrips, and Maruca vitrata Fabricius populations in cowpea fields. As reported by Araujo and Xavier (2009), the CNSL mainly contains anacardic acid, cardanol, cardol, 2-methylcardol in varying proportions and they are known to exhibit strong larvicidal activity against mosquitoes (Mukhopadhyay et al., 2010; Oliveira et al., 2011; Raraswati et al., 2014; Paiva et al., 2017). Similarly,

100% mortality of adults of *Sitophilus oryzae* L. (Coleoptera: Curculionidae) exposed to CNSL effects during five times (24h, 48h, 72h 96h, and 120h) was reported by Buxton *et al.* (2017). Moreover, Chinniah *et al.* (2020) reported that CNSL solution at 3% reduced 52% of the citrus leaf mite population, *Panonychus citri* McGregor.

As for the Neem oil, its effects of mite and insect pests have been reported by several authors (Fazil & Ansari, 2011; Oliveira et al., 2013; Traoré et al., 2019ab; Adango et al., 2020; Savi et al., 2021b; Azandémè-Hounmalon et al., 2022, in press). Furthermore, the Neem oil, Azadirachta indica, has insecticide and ovicidal potentials against insects and mites (Zongo et al., 1993; Ventura & Ito, 2000; Tedeschi et al., 2001; Schukla & Kumar, 2002; Viana & Prates, 2003; Bruce et al., 2004; Souza & Vendramim, 2005; Kumar et al., 2005; Haq, 2006; Venzon et al., 2007; Singha et al., 2007; Prabhat Kumar, 2007; Charbonneau et al., 2007; Yasmin et al., 2008; Faye, 2010; Fazil & Ansari, 2011; Oliveira et al., 2013; Traoré et al., 2019ab; Adango et al., 2020). The main compound of the Neem oil, Azadirachtin, widely admitted having fungicidal, insecticidal, and/or acaricidal properties (Karnavar, 1987; Bélanger & Musabyimana, 2005; Musabyimanan, 2005; Yarou et al., 2017; Traoré et al., 2019; Adango et al., 2020). Azadirachtin can be found in the leaves and the fruits; its toxicity is higher by ingestion than by contact, which allows its selective use against some phytophagous pests (Akol et al., 2002; Gaspari et al., 2007). Azadirachtin has an anti-appetite effect and can also acts by directly killing or repelling insects, affect egg-laying by females as well as molting and larval growth of certain arthropods (Stoll, 2002; Bélanger & Musabyimana, 2005; Guève et al., 2011; Gbedomon et al., 2012; Bernadi et al., 2013). In their study, Mouffok et al. (2008) reported the negative effects of azadirachtin on hormonal secretion, morphogenetic development, respiration of several insect species, and on the development of insect tissues (i.e., muscular, nervous, glandular, etc.). They also reported that azadirachtin is an anorexic agent since they are once in contact with it. Hence, the insect no longer eats and it finally dies. The high mortality of T. evansi (adults and eggs) recorded with Neem oil witnesses the sensitivity of all life stages of the mite pest to that azadirachtin. Our results showed that Acarius, the chemical insecticide, is not more effective than Neem oil and CNSL in controlling T. evansi.

Irrespective of its doses, Neem oil and the 4% or 5% solutions of CNSL, none of the eggs treated with those products survived and none of the survived females could oviposit. Thus, this suggests that *T. evansi* populations treated with Neem oil or CNSL at that concentration could seldom develop pesticide resistance, since no progeny could be expected and that could have survived to adulthood to initiate new populations (Osakabe *et al.*, 2009; Fortin *et al.*, 2012). In contrast, such risks are present with Acarius. Neem oil or

CNSL are derived from a vegetal, and thus, are expected to be less toxic to non-target organisms such as mammals, birds, aquatic fauna, and its use in biological or conventional agriculture (Mouffok *et al.*, 2008; Faye, 2010; Habou *et al.*, 2013). In addition, they are some relatively low-cost acaricide since the half of the recommended dose of Neem oil and a 4% solution of CNSL are enough to get maximal effect within 24 hours after application in the laboratory.

Conclusion

It appears from our results that Neem oil and CSNL could be considered for the integrated management (IPM) strategies against *T. evansi* in tomato fields. However, because our study was conducted in the laboratory, future studies are still needed to explore all the implications of using these biopesticides as alternatives to conventional synthetic acaricides against *T. evansi*, as well as their compatibility with biological control agents such as *Phytoseiulus longipes* Evans on tomato crop.

References:

- Adango, E., Onzo, A., & Daoudou, C.O.G.W. (2020). Evaluation de l'activité acaricide de quelques biopesticides sur l'acarien tarsonème, *Polyphagotarsonemus latus* Banks (Acari : Tarsonemidae) infestant l'aubergine gboma (*Solanum macrocarpon* L.) au Sud-Bénin. European Scientific Journal 16(15), 442-463. Doi:10.19044/esj.2020.v16n15p442.
- 2. Akol, A.M., Sithanantham, S., Njagi, P.G.N., Varela, A., & Mueke, J.M. (2002). Relative safety of sprays of two neem insecticides to *Diadegma mollipla* (Holmgren), a parasitoid of the diamondback moth: effects on adult longevity and foraging behavior. Crop Protection 21, 853-859.
- 3. Asogwa, E.U., Mokwunye, I.U., Yahaya, L.E., & Ajao, A.A. (2007). Evaluation of cashew nut shell liquid as a potential natural insecticide against termites (soldiers and worker's caste). Research Journal of Applied Sciences 2, 939-942.
- 4. Azandémè-Hounmalon, G.Y., Affognon, H.D., Assogba-Komlan, F., Tamó, M., Fiaboe, K.K.M., Kreiter, S., & Martin, T. (2015). Farmers' control practices against the invasive red spider mite, *Tetranychus evansi* Baker & Pritchard in Benin. Crop Protection 76, 53-58.
- Babu, A., Perumalsamy, K., Subramaniam, M.S.R., & Muraleedharan, N. (2008). Use of Neem kernel aqueous extract for the management of red spider mite infesting tea in South India. Journal of Plant Crops 36(3), 393-397.

- Bensoussan, N., Santamaria, M.E., Zhurov, V., Diaz, I., Grbić, M., & Grbić, V. (2016). Plant-herbivore interaction: Dissection of the cellular pattern of *Tetranychus urticae* feeding on the host plant. Frontiers in Plant Science 7, 1105.
- Bernardi, D., Botton, M., Cunha, U.S., Bernardi, O., Malausa, T., Garcia, M.S., & Nava, D.E. (2012). Effects of azadirachtin on *Tetranychus urticae* (Acari: Tetranychidae) and its compatibility with predatory mites (Acari: Phytoseiidae) on strawberry. Pest Management Science 69, 75-80. <u>https://doi.org/10.1002/ps.3364</u>.
- 8. Blair, B.W. (1983). *Tetranychus evansi* Baker and Pritchard (Acari; Tetranychidae), a new pest of tobacco in Zimbabwe. CORESTA. Phytopathology and Agronomy Study Group, Bergerac, France: 1-6.
- 9. Blair, B.W. (1989). Laboratory screening of acaricides against *Tetranyxchus evansi* Baker & Pritchard. Crop Protection 8, 217-222.
- 10. Bonato, O. (1999). The effect of temperature on life history parameters of *Tetranychus evansi* (Acari: Tetranychidae). Experimental and Applied Acarology 23, 11-19.
- Boubou, A., Migeon, A., Roderick, G., & Navajas, M. (2011). Recent emergence and worldwide spread of the red tomato spider mite, *Tetranychus evansi:* genetic variation and multiple cryptic invasions. Biological Invasions 13(1), 81-92.
- 12. Buxton, T., Takahashi, S., Niwata, I., Owusu, E.O., & Kim, C. (2017). Isolation and characterization of the insecticidal compounds in *Anacardium Occidentale* (cashew nut) shell liquid against the rice weevil, *Sitophilus oryzae* L. (Coleoptera: Curculionidae). Journal of Entomology and Zoology Studies 5(2), 1241-1246.
- 13. Charbonneau, C., Côté, R., & Charpentier, G. (2007). Effects of azadirachtin and of simpler epoxyalcohols on survival and behaviour of *Galleria mellonella* (Lepidoptera). Journal of Applied and Entomology 131, 447-452.
- Chaudhuri, T.C. (1999). Pesticide residues in tea. In: Jain NK, editor. Global advances in tea science. New Delhi (India): Aravali Books 882 p.
- 15. Chinniah, C., Naveena, K., & Shanthi, M. (2020). Evaluation of certain plant oils for their acaricidal property against Citrus leaf mite, *Panonychus citri* on acid lime. Journal of Pharmacognosy and Phytochemistry 9 (2), 2431-2434.
- Copping, L.G., & Menn, J.J. (2000). Biopesticides: a review of their action, applications and efficacy. Pest Management Science 56, 651-676.
- 17. Dimetry, N.Z., Amer, S.A.A., & Reda, A.S. (1993). Biological activity of two neem seed kernel extracts against the two-spotted spider mite

Tetranychus urticae Koch Journal of Applied and Entomology 116, 308-312.

- Dimetry, N.Z., Amer, S.A.A., & Reed, A.S. (1993). Biological activity of two neem seed kernel extracts against the two-spotted spider mite *Tetranychus urticae* Koch. Journal of Applied and Entomology 116, 308-312.
- <u>Djossou</u>, R., Azandémè-Hounmalon, G.Y., Onzo, A., Gnanvossou, D., Assogba-Komlan, F., & Tamò, M. (2021). Susceptibility of ten tomato cultivars to attack by *Tetranychus evansi* Baker & Pritchard (Acari: Tetranychidae) under laboratory conditions. International Journal of Tropical Insect Science 41 (1) <u>https://doi.org/10.1007/s42690-020-00221-9.</u>
- 20. Ferragut, F., & Escudero, L.A. (1999). *Tetranychus evansi* Baker & Pritchard (Acari, Tetranychidae) una nueva arana roja en los cultivos horticolas espanoles. <u>Boletín de Sanidad Vegetal, Plagas</u> 25, 157-164.
- 21. Gaspari, M., Lykouressis, D., Perdikis, D., & Polissiou, D. (2007). Nettle extract effects on the aphid *Myzus persicae* and its natural enemy, the predator *Macrolophus pygmaeus* (Hem: Miridae). Journal of Applied and Entomology 131, 652-657.
- 22. Golec, J.R., Hoge, B., & Walgenbach, J.F. (2020). Effect of biopesticides on different *Tetranychus urticae* Koch (Acari: Tetranychidae) life stages. Crop Protection 128, 105015 https://doi.org/10.1016/j.cropro.2019.10501.
- 23. Gotoh, T., Araki, R., Boubou, A., Migeon, A., Ferragut, F., Navajas, M. (2009). Evidence of co-specificity between *Tetranychus evansi* and *Tetranychus takafujii* (Acari: Prostigmata, Tetranychidae): comments on taxonomic and agricultural aspects. International Journal of Acarology 35, 485-501.
- 24. Gotoh, T., Fujiwara, S., & Kitashima, Y. (2010). Susceptibility to acaricides in nine strains of the tomato red spider mite *Tetranychus evansi* (Acari: Tetranychidae). International Journal of Acarology 37, 93-102.
- 25. Hazarika, L.K., Bhuyan, M., Hazarika, B.N. (2009). Insect pests of tea and their management. Annual Review of Entomology 54, 267-284.
- 26. Kennedy, G.G. (2003). Tomato, pests, parasitoids and predators: tritrophic interactions involving the genus Lycopersicon. Annual Review of Entomology 48, 51-72.
- 27. Kerala Agricultural University (2009). The adhoc package of practices recommendations for organique farming. Directorate of Research, Vellanikkara, KAU Post 680 656 Thrissur, Kerala, India.
- 28. Knapp, M., Wagener, B., & Navajas, M.J.A.E. (2003). Molecular discrimination between the spider mite *Tetranychus evansi* Baker &

Pritchard, an important pest of tomatoes in southern Africa, and the closely related species *T. urticae* Koch (Acarina: Tetranychidae). African Entomology 11, 300-304.

- 29. Knegt, B., Meijer, T.T., Kant, M.R., Kiers, T.E., Egas, M. (2019). *Tetranychus evansi* spider mite populations suppress tomato defenses to varying degrees. Ecology and Evolution 10, 4375-4390 DOI: 10.1002/ece3.6204.
- 30. Kumar, P., Poehling, H.M., & Borgemeister, C. (2005). Effects of different application methods of azadiracthin against sweetpotato whitefly *Bemisia tabaci* Gennadius (Hom., Aleyrodidae) on tomato plants. Journal and Applied Entomology 129, 489-497.
- 31. Kumral, N.A., Gencer, N.S., Susurluk, H.S., & Yalcin, C. (2011). A comparative evaluation of the susceptibility to insecticides and detoxifying enzyme activities in *Stethorus gilvifrons* (Coleoptera: Coccinellidae) and *Panonychus ulmi* (Acarina: Tetranychidae). International Journal of Acarology 37(3), 255-268.
- 32. Makundi, R.H., & Kashenge, S. (2002). Comparative efficacy of neem, Azadirachta indica, extract formulations and the synthetic acaricide, Amitraz (Mitac), against the two spotted spider mites, *Tetranychus urticae* (Acari: Tetranychidae), on tomatoes, *Lycopersicum esculentum*. Zeitschrift für Pflanzenkrankheiten und Pflanzenschutz/Journal of Plant Diseases and Protection 109, 57-63.
- 33. Marčić, D. & Medo, I. (2014). Acaricidal activity and sublethal effects of an oxymatrine based biopesticide on two-spotted spider mite (Acari: Tetranychidae). Experimental and Applied Acarology 64, 375-391. https://doi.org/10.1007/s10493-014-9831-x.
- 34. Migeon, A. (2005). Un nouvel acarien ravageur en France. *Tetranychus evansi* Baker and Pritchard. Phytoma 579, 38-42.
- 35. Migeon, A., & Dorkeld, F. (2018). Spider Mites Web: A comprehensive database for the Tetranychidae. Retrieved from http://www.montpellier.inra.fr/CBGP/spmweb.
- 36. Migeon, A., Ferragut, F., Escudero-Colomar, L., Fiaboe, K., Knapp, M., de Moraes, G., Ueckermann, E., & Navajas, M. (2009). Modelling the potential distribution of the invasive tomato red spider mite, *Tetranychus evansi* (Acari: Tetranychidae). Experimental and Applied Acarology 48, 199-212.
- Moghadam, M.M., Ghadamyari, M., & Talebi, K. (2012). Resistance mechanism to fenazaquin in Iranian populations of two spotted spider mite, *Tetranychus urticae* (Acari: Tetranychidae). International Journal of Acarology 38(2), 138-145.

- 38. Moraes, G.J., & McMurtry, J.A. (1986). Suitability of the spider mite *Tetranychus evansi* as prey for *Phytoseiulus persimilis*. Entomology and Experimental Applied 40, 109-115.
- 39. Mordue, A.J., & Nisbet, A.J. (2000). Azadirachtin from the neem tree *Azadirachta indica*: its action against insects. Anais da Sociedade Entomológica do Brasil 29, 615-632. https://doi.org/10.1590/S0301-80592000000400001.
- 40. Mouffok, B., Raffy, E., Urruty, N., & Zicola, J. (2008). Le neem, un insecticide biologique efficace. Département : Génie Biologique Projet Tutoré du S2, 16p.
- 41. Moutia, L.A. (1958). Contribution to the study of some phytophagous Acarina and their predators in Mauritius. Bulletin of Entomological Research 49, 59-75.
- 42. Mukhopadhyaya, A.K., Hatib, A.K., Tamizharasua, W., & Babu, P.S. (2010). Larvicidal properties of cashew nut shell liquid (*Anacardium occidentale* L) on immature stages of two mosquito species. Journal of Vector Borne Diseases 47, 257-260.
- 43. Muraleedharan, N., & Selvasundaram, R. (1996). UPASI hand book of tea culture. Coonoor (India): Sec., Published by the Secretary General, UPASI 23: 33p.
- 44. Musabyimana, T., Saxena, R.C.E., Kairu, W., Ogol, C.P.K., & Khan, Z.R. (2001). Effects of neem seed derivatives on behavioral and physiological responses of the *Cosmopolites sordidus* (Coleoptera: Curculionidae) Journal of Economic Entomology 94, 449-454.
- 45. Navajas, M., de Moraes, G.J., Auger, P., & Migeon, A. (2013). Review of the invasion of *Tetranychus evansi*: Biology, colonization pathways, potential expansion and prospects for biological control. Experimental and Applied Acarology 59, 43-65.
- 46. Nyoni, B.N., Gorman, K., Mzilahowa, T., Williamson, M.S., Navajas, M., Field, L.M., & Bass, C. (2011). Pyrethroid resistance in the tomato red spider mite, *Tetranychus evansi*, is associated with mutation of the para-type sodium channel. Pest Management Science 67, 891-7.
- 47. Oliveiraa, M.S.C., de Morais, S.M., Magalhães, D.V., Batistaa, W.P., Vieirac, Íc.G.P., Craveiroc, A.A., de Manezes, J.E.S.A., Carvalhoe, A.F., & de Lima, G.P.G. (2011). Antioxidant, larvicidal and antiacetyl-cholinesterase activities of cashew nut shell liquid constituents. Acta Tropica 117, 165-170.
- 48. Painuly, J.P., & Dev, S.M. (1998). Environmental dimensions of fertilizer and pesticide use: Relevance to Indian agriculture. International Journal of Environnment and Pollution 10(2), 273-288.
- 49. Paiva, D.R., de Lima, D.P., Avvari, N.P., de Arruda, E.J., Cabrini, I., Marques, M.R., dos Santos, E.A., Biaggio, F.C., Sangi, D.P., &

Beatriz, A. (2017). A potent larvicidal agent against *Aedes aegypti* mosquito from cardanol. Anais da Academia Brasileira de Ciências 89, 373-382.

- 50. Raraswati, G.R., Sudarsono, S., & Mulyaningsih, B. (2014). Activité larvicide d'un mélange de liquide de coque de noix de cajou et d'extrait hydrosoluble de fruit de noix de lavage (Sapindus rarak DC.) contre les larves de 3ème stade *d'Aedes aegypti*. Biologie Medecine Chimie des Produits Naturels, 3 (2), 53-57.
- 51. RECA (2013). Réseau National des Chambres d'Agriculture du Niger ; Fiche conseil pour la matière active : Abamectine (acaricide insecticide) 5p.
- 52. Reddy, P.P. (2016). Selective pesticides in IPM. In: Sustainable crop protection under protected cultivation. Springer, Singapore p. 121-131.
- 53. Roobakkumar, A., Subramaniam, M.S.R., Babu, A., & Muraleedharan, N. (2010). Bioefficacy of certain plant extracts against the red spider mite, *Oligonychus coffeae* Nietner (Acarina: Tetranychidae) infesting tea. International Journal of Acarology 36(3), 255-258.
- 54. Savi, P.J., de Moraes, G.J., Melville, C.C., & Andrade, D.J. (2019a). Population performance of *Tetranychus evansi* Baker & Pritchard (Acari: Tetranychidae) on African tomato varieties and wild tomato genotypes. Experimental Applied and Acarology 77, 555-570. https://doi.org/10.1007/s10493-019-00364-6.
- 55. Savi, P.J., Martins, M.B., de Moraes, G.J., Hountondji, F.C.C., & de Andrade, D.J. (2021b). Bioactivity of oxymatrine and azadirachtin against *Tetranychus evansi* (Acari: Tetranychidae) and their compatibility with the predator *Phytoseiulus longipes* (Acari: Phytoseiidae) on tomato. Systematic Applied and Acarology 26(7), 1264-1279 https://doi.org/10.11158/saa.26.7.7.
- 56. Sarwar, M. (2015). Biopesticides: une ligne d'action efficace et respectueuse de l'environnement pour inhiber les insectes nuisibles. International Journal of Engineer Advan Research Technology 1(2),10-15.
- 57. Singha, A., Thareja, V., & Singla, A.K. (2007). Application of neem seed kernel extracts result in mouthpart deformites and subsequent mortality in *Nezara viridula* (L.) (Hem: Pentatomidae). Journal of Applied and Entomology 131, 197-201.
- 58. Soto, A., Venzon, M., Oliveira, R.M., Oliveira, H.G., & Pallini, A. (2010). Alternative Control of *Tetranychus evansi* Baker & Pritchard (Acari: Tetranychidae) on Tomato Plants grown in Greenhouses. Neotropical Entomology 39(4), 638-644.

- Souza, A.P., & Vendramim, J.D. (2005). Translaminar, sistemic and topical effect of aqueous extract of neem seed on *Bemisia tabaci* (Genn.) biotype B on tomato plants. Neotropical Entomology 34, 83-87.
- 60. Sundaram, K.M.S., & Sloane, L. (1995). Effects of pure and formulated Azadirachtin, a neem-based biopesticide on the phytophagous spider mite *Tetranychus urticae*. Journal of Environnment. Science Health B 30, 801-814.
- 61. Tedeschi, R., Alma, A., & Tavella, L. (2001). Side-effects of three neem (*Azadirachta indica* A. Juss) products on the predator *Macrolophus caliginosus* Wagner (Het., Miridae). Journal of Applied and Entomology 125, 397-402.
- 62. Toroitich, F.J., Knapp, M., Nderitu, J.H., Olubayo, F.M., & Obonyo, M. (2014). Susceptibility of geographically isolated populations of the Tomato red spider mite (*Tetranychus evansi* Baker & Pritchard) to commonly used acaricides on tomato crops in Kenya. Journal of Entomology and Acarology Research 46, 18-23.
- 63. Tsagkarakou, A., Cross-Ateil, S., & Navajas, M. (2007). First record of the invasive mite *Tetranychus evansi* in Greece. Phytoparasitica 35, 519-522.
- Tsagkarakou, A., Pasteur, N., Cuany, A., Chevillon, C., & Navajas, M. (2002). Mechanisms of resistance to organophosphates in *Tetranychus urticae* (Acari: Tetranychidae) from Greece. Insect Biochemistry and Molecular 32, 417-424.
- Vasanthakumar, D., Babu, A., Shanmugapriyan, R., & Subramaniam, S.R. (2013). Impact of Azter (azadirachtin 0.15% EC), a neem-based pesticide, against tea red spider mite, *Oligonychus coffeae* Neitner (Acarina: Tetranychidae), and its natural enemies, International Journal of Acarology 39(2), 140-145. DOI: 10.1080/01647954.2012.754494.
- 66. Ventura, M.U., & Ito, M. (2000). Antifeedant activity of Melia azedarach (L.) extracts to *Diabrotica speciosa* (Genn.) (Coleoptera: Chrysomelidae) beetles. Brazilian Archives of Biology and Technology 43, 215-219.
- 67. Venzon, M., Rosado, M.C., Fadini, M.A.M., Ciociola, Jr.A.I., & Pallini, A. (2005). The potential of NeemAzal for the control of coffee leaf pests. Crop Protection 24, 213-219.
- 68. Venzon, M., Rosado, M.C., Pallini, A., Fialho, A., & Pereira, C.J. (2007). Lethal and sublethal toxicity of neem on green peach aphid and on its predator Eriopis connexa Pesquisa Agropecuária Brasileira 42, 627-631.

- 69. Viana, P.A., & Prates, H.T. (2003). Desenvolvimento e mortalidade larval de *Spodoptera frugiperda* em folhas de milho tratadas com extrato aquoso de folhas de *Azadirachta indica* Bragantia 62, 69-74.
- 70. Yasmin, N., Khan, M.F., Channa, M.S., & Zeeshan, A. (2008). Effects of a neem sample on protein patterns of *Bactrocera cucurbitae*. Turkish Journal of Zoology 32, 1-5.